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**1. PURPOSE**

This Standard Operating Procedure (SOP) describes methods for anesthetizing mice.

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**2. RESPONSIBILITY**

Principal Investigators (PIs) and their research staff, veterinary care staff.

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**3. INTRODUCTION**

- 3.1. Mice are not routinely fasted prior to anesthesia due to their inability to vomit.
- 3.2. Rodents can be anesthetized with either inhalant gas or injectable drugs. The use of inhalant gases is the preferred method of anesthesia whenever possible.
- 3.3. Heat loss is rapid in anesthetized rodents. Keep animals warm by covering them (e.g. gauze pad or towel) and/or providing a heat source until the animal has recovered from anesthesia.
- 3.4. Never leave an anesthetized animal unattended.

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**4. MATERIALS**

- 4.1. Material or equipment to provide or conserve body heat (e.g. gauze pads, heating disc or pad, warm-water circulating pad)
- 4.2. Ophthalmic ointment (natural tears)
- 4.3. Gas anesthesia machine (calibrated within the last 12 months) with adequate gas scavenging system or filter
- 4.4. Induction chamber constructed of a see-through material (glass, polycarbonate, etc.)
- 4.5. Isoflurane
- 4.6. Ketamine (100mg/mL) \*Controlled Drug
- 4.7. Xylazine (20mg/mL)
- 4.8. Acepromazine (10mg/mL)
- 4.9. Atipamezole 5 mg/ml
- 4.10. 2,2,2-Tribromoethanol (Avertin)
- 4.11. Tertiary amyl alcohol
- 4.12. Sterile isotonic saline (0.9% saline) or sterile water for injection
- 4.13. Crushed ice or ice pack

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**5. PROCEDURES FOR ADULT MICE**

- 5.1. Isoflurane anesthesia:
  - 5.1.1. Induction:
    - 5.1.1.1. Place the animal in the induction chamber
    - 5.1.1.2. Adjust the oxygen flowmeter to 0.8 to 1.5 L/min.
    - 5.1.1.3. Adjust the isoflurane vaporizer to 3% to 5%. .

- 5.1.2. Maintenance:
  - 5.1.2.1. Use the mask connected to the Bain circuit,
  - 5.1.2.2. Adjust the flowmeter to 0.4 to 0.8 L/min.
  - 5.1.2.3. Adjust the isoflurane vaporizer to 2 to 2.5%.
  - 5.1.2.4. Apply ophthalmic ointment (natural tears) to both eyes to prevent dryness and damage to the cornea. Reapply as needed.
- 5.1.3. Recovery:
  - 5.1.3.1. Turn off the isoflurane vaporizer but keep the animal on oxygen.
  - 5.1.3.2. Transfer animal to their cage once it begins to move and allow to recover fully (sternal position).
- 5.2. Ketamine/Xylazine/Acepromazine anesthesia:
  - 5.2.1. Injectable anesthetic dose can vary with the sex, the age, the strain, and the body condition of the animal.
  - 5.2.2. Contact your veterinarian for advice on the appropriate dose prior to use.
  - 5.2.3. Anesthetic dose: ketamine 100mg/kg, xylazine 10mg/kg, acepromazine 3mg/kg.
  - 5.2.4. Prepare the solution the day before or shake it thoroughly before use.
  - 5.2.5. To prepare cocktail, in a sterile vial or bottle with a rubber stopper, mix:
    - 1mL of ketamine (100mg/mL)
    - 0.5mL xylazine (20mg/mL)
    - 0.3mL acepromazine (10mg/mL)
    - 8.2mL of sterile isotonic saline or sterile water for injection.
  - 5.2.6. Label as "Mouse Cocktail" and indicate expiration date on vial or bottle (maximum 6 months).
  - 5.2.7. Mixed cocktail should be protected from light and stored in a cool place.
  - 5.2.8. Administer 0.05-0.1mL/10g body weight intraperitoneally.
  - 5.2.9. Apply ophthalmic ointment (natural tears) to both eyes to prevent dryness and damage to the cornea. Reapply as needed.
  - 5.2.10. Duration of anesthesia is approximately 20 minutes.
  - 5.2.11. After 20 minutes, a half dose may be administered as needed.
  - 5.2.12. Administer atipamezole 1-2 mg/kg SC or IP to improve respiration or speed up the recovery if needed. Atipamezole is the antidote for xylazine.
- 5.3. 2,2,2-Tribromoethanol (TBE or Avertin) anesthesia:
  - 5.3.1. Anesthetic dose is 250 to 500mg/kg.
  - 5.3.2. Avertin administration can result in sensitization of the animal; thus, it is recommended to be given only on a single occasion.
  - 5.3.3. Avertin stock solution:
    - 5.3.3.1. In a sterile container add 25g of 2,2,2-tribromoethanol to 15.5mL tertiary amyl alcohol and dissolve by heating to 50°C and stirring until completely dissolved.
    - 5.3.3.2. Store protected from light (wrapped in foil or in an amber container) in the refrigerator or freezer.
  - 5.3.4. Label as "Avertin Stock Solution" and indicate expiration date (up to 1 year).
  - 5.3.5. Avertin working solution:
    - 5.3.5.1. In a sterile container, mix 0.5mL of the stock solution in 39.5mL of sterile isotonic saline.
    - 5.3.5.2. Heat solution to 40°C then shake well until completely dissolved. Mixture should be clear.
    - 5.3.5.3. Filter the working solution through 0.2 micron filter.

- 5.3.5.4. Store protected from light (wrapped in foil or in an amber container) at 4°C.
- 5.3.5.5. Label as "Avertin 20mg/mL" and indicate expiration date (maximum 4 months)
- 5.3.6. Inject 0.1-0.25mL of working solution/10g body weight, intraperitoneally.
- 5.3.7. Apply ophthalmic ointment (natural tears) to both eyes to prevent dryness and damage to the cornea. Reapply as needed.
- 5.3.8. Duration of anesthesia is approximately 20 minutes.

## 6. PROCEDURES FOR NEONATAL MICE

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### 6.1. Hypothermia:

- 6.1.1. Use only in animals less than 7 days of age.
- 6.1.2. Provides immobilization and mild analgesia for short, minor procedures.
- 6.1.3. Protect pup in a glove or paper-lined tube to avoid damage to the skin.
- 6.1.4. Induction:
  - 6.1.4.1. Immerse pup in ice water or crushed ice for 3 to 4 minutes.
- 6.1.5. Maintenance:
  - 6.1.5.2. Place pup on a paper-covered ice pack.
  - 6.1.5.3. Use a fiber optic surgical lamp if necessary as incandescent lamps will warm the animal and interfere with anesthesia.
  - 6.1.5.4. Duration of anesthesia is approximately 10 minutes.
- 6.1.6. Recovery:
  - 6.1.6.1. Remove animal from ice pack and allow to warm.
  - 6.1.6.2. Recovery time can be up to 1 hour.

### 6.2. Isoflurane anesthesia:

- 6.2.1. Neonates require higher concentration of isoflurane than adults (maintenance at 3-4%). See section 5.1 for detailed procedure.

## SOP REVISION HISTORY

DATE	PREVIOUS VERSION	NEW VERSION
2016.01.15	(NO TEXT)	<b>Addition of Rodent Procedure Log (annex)</b>
2016.03.16	5.3.1 Anesthetic dose is 240mg/kg.	5.3.1 Anesthetic dose is <del>240</del> <b>250 to 500</b> mg/kg.
2016.03.16	5.2.1 and 5.2.2 (NO TEXT)	<b>5.2.1 Injectable anesthetic dose can vary with the sex, the age, the strain, and the body condition of the animal.</b> <b>5.2.2 Contact your veterinarian for advice on the appropriate dose prior to use.</b>
2016.03.16	5.2.12 (NO TEXT)	<b>5.2.12 Administer atipamezole 1-2 mg/kg SC or IP to improve respiration or speed up the recovery if needed. Atipamezole is the antidote for xylazine.</b>

Investigator:	Protocol:
Procedure:	Performed by:

**Instructions:** complete this log for rodent procedures requiring anesthesia, analgesia or post-procedure care (ex. surgeries, experimental infection). Keep the log in the housing room while active and in your files for 3 years for future review by the Quality Assistant and/or the FACC.

**ANALGESIA**

- carprofen: mouse: 20mg/kg, rat: 5-10 mg/kg, SC, every 24 hrs
- buprenorphine: mouse: 0.1mg/kg SC or IP every 4-8 hrs;  
rat: 0.05mg/kg, SC or IP, every 8-12 hrs
- lidocaine/bupivacaine (local analgesic)
- other: \_\_\_\_\_

**ANESTHESIA**

- isoflurane 2-2.5%
- ketamine/xylazine/acepromazine\*:  
mouse: 100 mg/kg (K)- 10 mg/kg (X)- 3 mg/kg (A) IP  
rat: 50 mg/kg (K)- 5 mg/kg (X)- 1 mg/kg (A); IP or IM
- other: \_\_\_\_\_

**OTHER AGENTS ADMINISTERED**

- \_\_\_\_\_
- \_\_\_\_\_
- \_\_\_\_\_

Animal ID	Date	Anesthesia		Analgesia		Other		Heat Source Provided		Recovery time	Comments/observations	Initials
		dose	time	dose	time	dose	time	procedure	recovery			
1								<input type="checkbox"/>	<input type="checkbox"/>			
2								<input type="checkbox"/>	<input type="checkbox"/>			
3								<input type="checkbox"/>	<input type="checkbox"/>			
4								<input type="checkbox"/>	<input type="checkbox"/>			
5								<input type="checkbox"/>	<input type="checkbox"/>			
6								<input type="checkbox"/>	<input type="checkbox"/>			
7								<input type="checkbox"/>	<input type="checkbox"/>			
8								<input type="checkbox"/>	<input type="checkbox"/>			
9								<input type="checkbox"/>	<input type="checkbox"/>			
10								<input type="checkbox"/>	<input type="checkbox"/>			
11								<input type="checkbox"/>	<input type="checkbox"/>			
12								<input type="checkbox"/>	<input type="checkbox"/>			
13								<input type="checkbox"/>	<input type="checkbox"/>			
14								<input type="checkbox"/>	<input type="checkbox"/>			

Comments/footnotes:

\*Dose can vary with the sex, the age, the strain, and the body condition of the animal.

## ANALGESIA

- carprofen: mouse: 20mg/kg, rat: 5-10 mg/kg, SC, every 24 hrs
- buprenorphine: mouse: 0.1mg/kg SC or IP every 4-8 hrs; rat: 0.05mg/kg, SC or IP, every 8-12 hrs
- OTHER \_\_\_\_\_

Initial the appropriate boxes when completed

	Animal ID	Date	Analgesia			SC fluids			Wet food			Time			Remove Sutures (Day 7-10)
			Day 1	Day 2	Day 3	Day 1	Day 2	Day 3	Day 1	Day 2	Day 3	Day 1	Day 2	Day 3	
1															
2															
3															
4															
5															
6															
7															
8															
9															
10															
11															
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14															

Comments/footnotes: